PHMB

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DATA EVALUATION RECORD

STUDY TYPE:

In Vitro Dermal Penetration Test

OPPTS 870,7600

DP BARCODE:

D261221

**SUBMISSION CODE:** S569727

P.C. CODE: 111801

**CASE TYPE:** Reregistration

TEST MATERIAL (PURITY):

PHMB (20.2%) and [14C]-PHMB (Specific activity:185GBq/732mg, the purity of

radio-labeled test substances has not been reported)

**CITATION**: Clowes, H.M. 1996. PHMB: In Vitro Absorption Through Human Epidermis.

September 6, 1996. Report No. CTL/P/5120. Prepared by Zeneca Central Toxicology Laboratory, Cheshire, UK. [MRID 44119301]. Unpublished Report.

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#### **EXECUTIVE SUMMARY:**

To evaluate the potential of penetration of PHMB through human skin, the absorption of PHMB was measured *in vitro* through human epidermis. A flow-through skin on chamber design was used and three aqueous PHMB concentrations (nominal concentrations of 200g /l, 20g/l and 2g/l) were applied. These dilutions were applied to the epidermal membranes at a rate of 10μl/cm<sup>2</sup>; the 200g/l concentration was also applied at 200μl /cm<sup>2</sup>. The 10μl/cm<sup>2</sup> applications were left unoccluded throughout the entire exposure period and the 200 μl/cm<sup>2</sup> was occluded.

The *in vitro* results indicated the absorption of PHMB from the nominal 20% solution through human epidermis was slow and linear over the 96h exposure period. The permeability coefficient (Kp) has been calculated to be  $5.47 \times 10^{-7}$  cm/h from the steady state rate of absorption (0-24h). Approximately 13 times more was absorbed from the  $200\mu l/cm^2$  occluded application ( $0.088\mu g/cm^2/h$ ) than from the  $10\mu l/cm^2$  unoccluded application ( $0.007\mu g/cm^2/h$ ).

The absorption rate of PHMB was very slow from the nominal 2% solution  $(0.005\mu g/cm^2/h)$  during the first 24h and slowed down further between 24 and 96h  $(0.003\mu g/cm^2/h)$ . Absorption from the nominal 0.2% solution was measured at  $0.001\mu g/cm^2/h$  during the first 24h and was below the limit of quantitation over the 24-96h period.

This study indicates a slow rate and extent of absorption of PHMB through human epidermis. However, this system did not take into account other skin structures such as dermis which may have an influence on PHMB absorption. In addition, these data do not account for the influence of systemic distribution and excretion kinetics upon dermal absorption of PHMB. Thus, this study is classified as **unacceptable** and does not satisfy the OPPTS guideline for dermal penetration study (OPPTS 870.7600).

**COMPLIANCE:** Signed and dated Quality Assurance, Good Laboratory Practice, Data Confidentiality, and Flagging statements were included.

### I. MATERIALS AND METHODS

### A. TEST SUBSTANCES

#### A.1 PHMB

Chemical name: Polyhexamethylene biguanide (PHMB)

PHMB content: 20.2%

Source: Zeneca Inc. Dighton, Mass., USA

Color: Faint Yellow Physical state: Liquid

Storage conditions: Ambient temperature and in the dark

# A.2 [14C] - PHMB

Source: Cambridge Research Biochemicals, Northwich, Cheshire, UK

Physical state: Faint yellow liquid

Radiochemical reference number: 4602 Specific activity: 1.85GBq/732mc,

Storage conditions: Freezer

# \* denotes the position of <sup>14</sup>C-labelled atoms

n = 1 - 40

Where 
$$X = -CH_2.CH_2.CH_2-NH_2^+.HCI$$

or  $-CH_2.CH_2.CH_2-NH-C-NH.CN$ 
 $NH$ 

or  $-CH_2.CH_2.CH_2-NH-C-NH_2^+.HCI$ 
 $NH$ 
 $NH$ 

#### A.2.1 Structure of the radiolabelled test substance

# B. EXPERIMENTAL PROCEDURES

## B.1 Liquid scintillation counting (LSC) conditions

Sample volume: 250µl

Counting period: 4 minutes or until 40000 counts had been registered

Scintillation fluid: Optiphase 'HISAFE' (10ml)

The limit of quantitation using the above procedure was set at between 0.002 and

0.2μg/ml depending on the application.

# B.2 Preparation of epidermal membranes

Extraneous tissue was removed from human whole skin samples. No information was given regarding which part of the body where the skin was taken from. The skin samples were immersed in water at 60°C for 40-45 seconds and the epidermis teased off the dermis. Each epidermal membrane was given an identifying number and stored frozen on aluminium foil until required for use.

# B.3 Measurement of membrane integrity

Samples of epidermis were mounted in glass diffusion cells with an exposed area of  $2.54 \text{cm}^2$ . The cells were placed in a water bath maintained at  $30 \pm 1^{\circ}\text{C}$ . The integrity of the membranes was determined by measurement of their electrical resistance across the skin membrane. Membranes with a calculated resistance smaller than  $10 \text{ k} \square$  were regarded as being lower than normal and not used for exposure to the test material.

# B.4 Measurement of test substance absorption

A flow-through skin on chamber design was used (See Figure 1). The receptor chambers of the cells were filled with a recorded volume of receptor fluid (distilled water) and placed in a water bath maintained at  $30 \pm 1^{\circ}$ C. A pre-treatment sample (0.25ml) was taken from each receptor chamber for analysis by LSC. An equal volume of fresh receptor fluid was added to each receptor chamber to replace the volume removed.

PHMB was applied to the skin membranes as dilutions in water at the dose rates indicated in **Table 1**. PHMB solutions were prepared by mixing cold PHMB with [ $^{14}$ C]-PHMB and distilled water to give the required concentrations. The [ $^{14}$ C]-PHMB was added in amounts to give activities ranging from approximately 3 x 10 $^6$  dpm/ml for the 0.2% dose to approximately 4 x 10 $^9$  dpm/ml for the 20% dose to maximize the analytical sensitivity of the varying dosing regimes. The  $10\mu$ l/cm $^2$  applications were left unoccluded and the  $200\mu$ l/cm $^2$  occluded (20% solution) for the duration of the exposure period (96h). These application doses were designed to simulate potential dermal exposure to PHMB during normal use.

At recorded intervals, samples (0.25ml) of the receptor fluid were taken for analysis by LSC. The volume of fluid in the receptor chamber was maintained by the addition of (0.25ml) of fresh receptor fluid to the chamber immediately after the removal of each sample.

#### II. RESULTS AND DISCUSSION

The results obtained in this study are summarized in **Tables 1 and 2**, where data are presented both in terms of absorption rate and in terms of amount and percent of the dose applied at 6, 8, 24 and 96h after exposure.

Absorption of PHMB from the nominal 20% solution through human epidermis was slow and linear over the 96h exposure period. The permeability coefficient (Kp) has been calculated to be  $5.47 \times 10^{-7}$  cm/h from the steady state rate of absorption (0-24h). Approximately 13 times more was absorbed from the  $200\mu l/cm^2$  occluded application (0.088 $\mu g/cm^2$ /h) than from the  $10\mu l/cm^2$  unoccluded application (0.007 $\mu g/cm^2$ /h).

The absorption rate of PHMB was very slow from the nominal 2% solution  $(0.005\mu g/cm^2/h)$  during the first 24h and slowed down further between 24 and 96h  $(0.003\mu g/cm^2/h)$ . Absorption from the nominal 0.2% solution was measured at  $0.001\mu g/cm^2/h$  during the first 24h and was below the limit of quantitation over the 24-96h period.

For the 20, 2 and 0.2% solutions from the  $10\mu$ l/cm<sup>2</sup> unoccluded applications, absorption was slow and linear over the initial 24 hours. The steady state rates of absorption are tabulated below:

#### III. SUMMARY AND CONCLUSION

This study indicates a slow rate and extent of absorption of PHMB through human epidermis. However, this system did not take into account other skin structures such as dermis which may have an influence on PHMB absorption. In addition, these data do not account for the influence of systemic distribution and excretion kinetics upon dermal absorption of PHMB. Thus, this study is classified as **unacceptable** and does not satisfy the OPPTS guideline for dermal penetration study (OPPTS 870.7600).

TABLE 1. MEAN PHMB ABSORPTION RATE THROUGH HUMAN EPIDERMIS <sup>a</sup>									
Concentra tion (g/L)	Actual Concentra tion (g/L)	Applied Volume (μL/cm	Applied Dose (µg/cm²	Number of Samples	Time Period (hour)	Absorption Rate (μg/cm²/hour ± SEM )			
					0 - 24	$0.001 \pm < 0.001$			
	0.000		000000		24 - 96	$<0.001 \pm <0.001$			
2	1.93	10 <sup>b</sup>	20	8	0 - 96	< 0.001 ± < 0.001			
					0 - 24	$0.005 \pm < 0.044$			
					24 - 96	$0.003 \pm < 0.001$			
20	20.9	10 <sup>b</sup>	200	8	0 - 96	$0.003 \pm < 0.001$			
					0 - 24	$0.009 \pm 0.003$			
		72			24 - 96	$0.007 \pm 0.002$			
200	197	10 <sup>b</sup>	2000	5	0 - 96	$0.007 \pm 0.002$			
					0 - 24	$0.110 \pm 0.044$			
					24 - 96	$0.083 \pm 0.029$			
200	201	200 <sup>b</sup>	402000	4	0 - 96	$0.088 \pm 0.033$			

Note: a. Taken From Page 19 Table 1. The  $10\mu$ /cm<sup>2</sup> applications were unoccluded throughout the entire period and the 200  $\mu$ /cm<sup>2</sup> was occluded.

TABLE 2. MEAN PHMB ABSORBED THROUGH HUMAN EPIDERMIS <sup>a</sup>									
Concentra tion (g/L)	Actual Concentra tion (g/L)	Applied Volume (µL/cm	Applied Dose (µg/cm <sup>2</sup>	Number of Samples	Expos ure Time (hour s)	Amount Absorbed (µg/cm²)	Percent Absorbed (%)		
					6	0.007	0.036		
					8	0.008	0.041		
****	V. (1997) 128-275.				24	0.017	0.088		
2	1.93	10 <sup>b</sup>	20	8	96	0.025	0.129		
					6	0.032	0.015		
					8	0.043	0.020		
		100			24	0.125	0.059		
20	20.9	10 <sup>b</sup>	200	8	96	0.307	0.146		
					6	0.123	0.006		
					8	0.141	0.007		
					24	0.237	0.012		
200	197	10 <sup>b</sup>	2000	5	96	0.708	0.036		
					6	0.784	0.001		
		li e			8	1.04	0.002		
				200	24	2.64	0.006		
200	201	200 <sup>b</sup>	402000	4	96	8.47	0.021		

Note: a. Taken From Page 19 Table 1.
 b. The 10μl/cm² applications were unoccluded throughout the entire period and the 200 μl/cm² was occluded.

PHMB

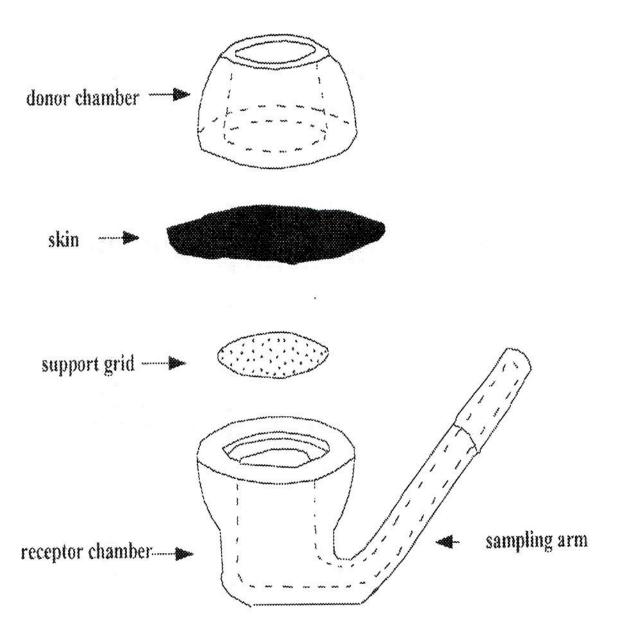


Figure 1. Diffusion Cell